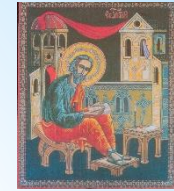




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Почетен член на "Съвета на Европейската научна и културна общност" (ЕСНКО)



# New aspects in the development of the interactive system for education in modelling and control of bioprocesses

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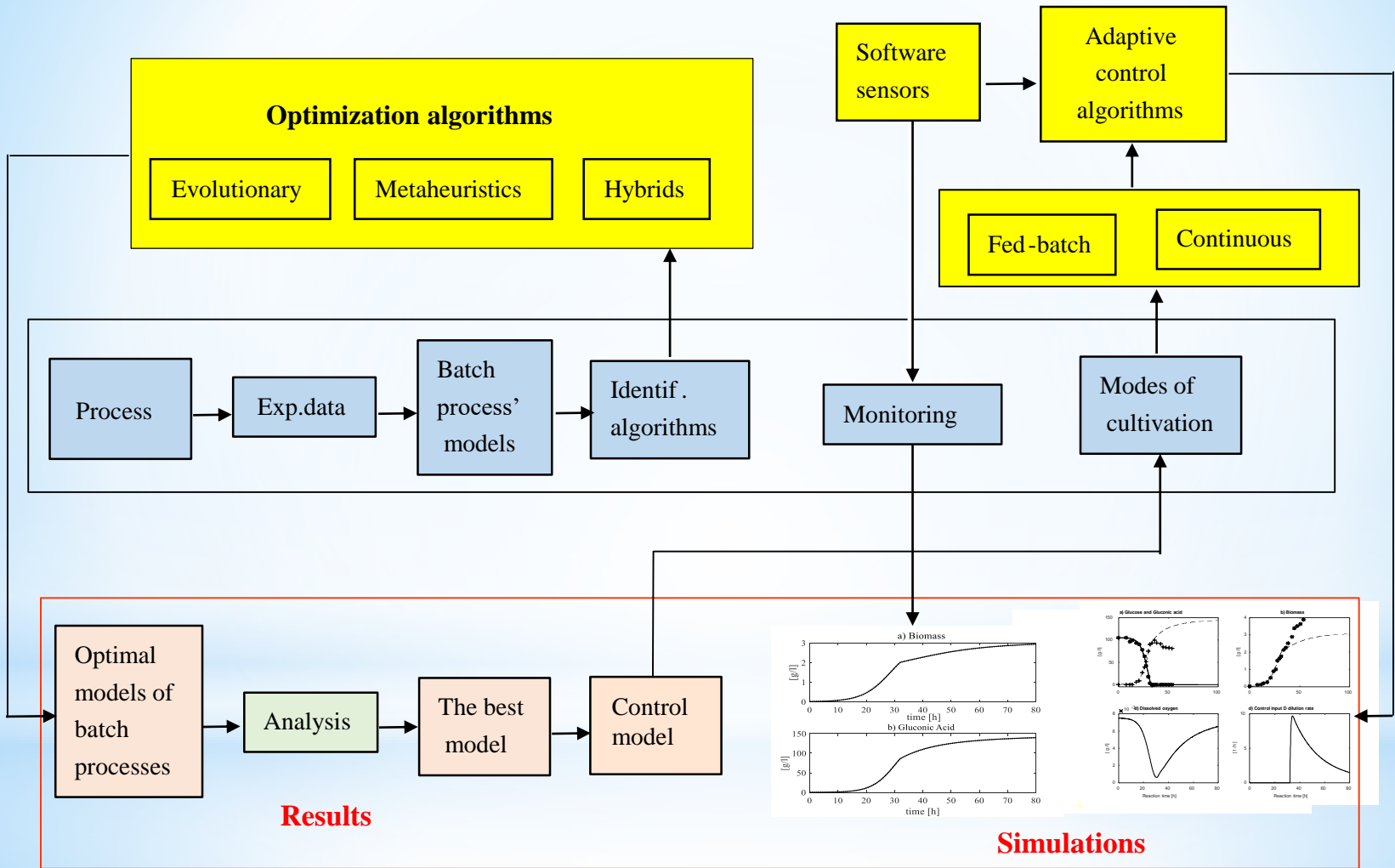
May 2023

Sibiu, Roumania

# Presentation Outline

- 1. General scheme of Open source system InSEMCoBio**
- 2. Evolutionary Algorithm Implemented in InSEMCoBio**
- 3. Multistep Model for the case of extracellular production of bacterial phytase at fed-batch cultivation of *E. coli***
- 4. Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen**
- 5. Adaptive Control of Protein Production Bioprocess with Three Physiological States**

# Open source system InSEMCoBio



# Evolutionary Algorithm Implemented in the Interactive System for Education in Modelling of Bioprocesses

The figure displays two screenshots of the InSEMCoBio software interface, showing the configuration of a fermentation process model and the selection of a metaheuristic algorithm.

**Figure 1a (Top):** Shows the "Identification Panel" with the following settings:

- Current Step:** Select Fermentation Process, Select Model and Kinetics, Load Experimental Data, Model Parameter Identification.
- Choose Fermentation Process:** E. coli MC4110 Fed-batch
- Choose Model and Kinetics:**
  - Mass Balance Equations:**   $dX/dt = \mu X - F/V \cdot X$ ,   $dS/dt = -1/Y_{xs} \mu X + (S_0 - S) \cdot F/V$ ,   $dO_2/dt = 1/Y_{ox} \mu X + K_{la} (O_2^* - O_2) - F/V \cdot O_2$ ,   $dV/dt = F$
  - Kinetic Models:**  Monod,  Contoa,  Andrew
- Buttons:** Set Model, Load Data
- Logs:** Table with columns Step and Record. Row: FP, E. coli MC4110 Fed-batch.

**Figure 1b (Bottom):** Shows the "Identification Panel" with the following settings:

- Current Step:** Select Fermentation Process, Select Model and Kinetics, Load Experimental Data, Model Parameter Identification.
- Choose Fermentation Process:** E. coli MC4110 Fed-batch
- Choose Model and Kinetics:**
  - Mass Balance Equations:**   $dX/dt = \mu X - F/V \cdot X$ ,   $dS/dt = -1/Y_{xs} \mu X + (S_0 - S) \cdot F/V$ ,   $dO_2/dt = 1/Y_{ox} \mu X + K_{la} (O_2^* - O_2) - F/V \cdot O_2$ ,   $dV/dt = F$
  - Kinetic Models:**  Monod,  Contoa,  Andrew
- Buttons:** Set Model, Load Data
- Choose Algorithm:** Evolutionary Algorithm
- Set Algorithm Parameters:** Max Iter: 10 (1, 100), Step: 0.5 (0, 5)
- Set Problem Parameters:** mumax: 0.45 (0.45, 0.52), ks: 0.01 (0.01, 0.05), k1: 1.8 (1.8, 2.05)
- Buttons:** Run, Plot Results
- Logs:** Table with columns MKA and Results. Row: EcoliFB, Monod, EA, Best solution:  $\mu = 0.46$   $k_s = 0.01$

Fig. 1a, b. Setting up a fermentation process model and metaheuristic algorithm parameters in InSEMCoBio

## Mathematical Model

The application of the general state space dynamical model to the fed-batch cultivation process of bacteria *E. coli* leads to the following nonlinear differential equation system :

$$\frac{dX}{dt} = \frac{\mu_{\max} S}{k_S + S} X + \frac{F}{V} X$$

$$\frac{dS}{dt} = -\frac{1}{Y_{XS}} \frac{\mu_{\max} S}{k_S + S} X + \frac{F}{V} (S_{in} - S)$$

$$\frac{dV}{dt} = F$$

$X$ – the concentration of the biomass, [g/L],

$S$ – the concentration of the substrate (glucose), [g/L];

$F$ – the feeding rate, [L/h];

$V$ – the volume of the bioreactor, [L];

$S_{in}$ – the initial glucose concentration in the feeding solution, [g/L];

$\mu$ – the specific growth rate described by Monod kinetics, [1/h];

$\mu_{\max}$ – the maximum growth rate, [1/h];

$k_S$ – a saturation constant, [g/L];

$Y_{XS}$ – a yield coefficient, [-].

# Experimental Data and results

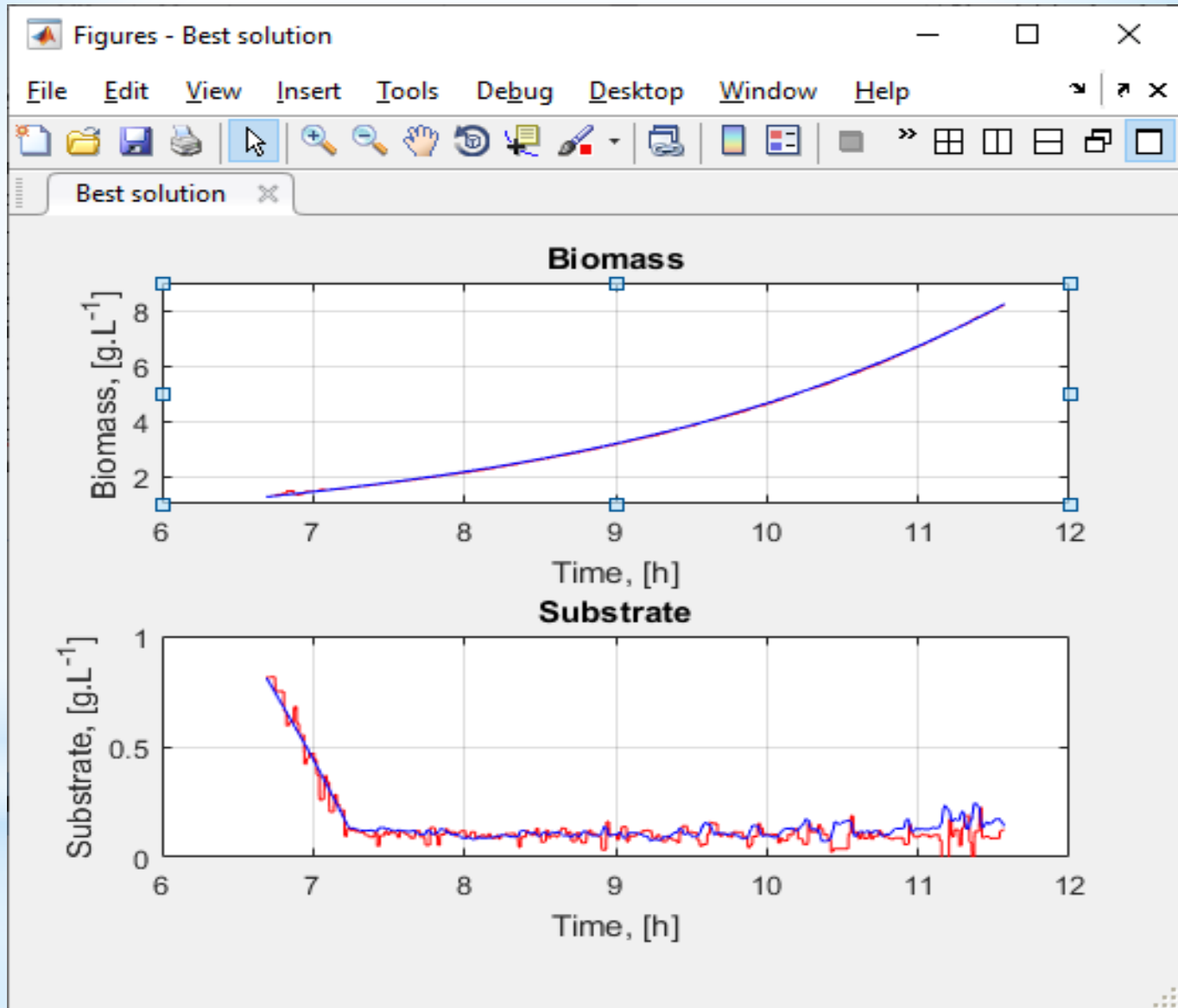


Fig. 1c Visualizing the results of the identification procedure in InSEMCoBio

# Multistep Modeling of a Class Bioprocesses

## Oxidative-fermentative growth model on glucose and oxidative on acetate

### Oxidative-fermentative growth on glucose

$$R_{ac} > 0 \quad \Rightarrow \quad \frac{d}{dt} \begin{bmatrix} X \\ S \\ A \\ P \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & -k_2 \\ 0 & k_3 \\ k_5 & k_6 \end{bmatrix} \begin{bmatrix} \mu_1 \\ \mu_2 \end{bmatrix} - \frac{F}{V} \begin{bmatrix} X \\ S - S_{in} \\ A \\ P \end{bmatrix}$$

$$\mu_1 = q_{s,crit} / k_1$$

$$\mu_2 = (q_s - q_{s,crit}) / k_2$$

$$q_{s,crit} = \frac{q_{o,max}}{k_{os}} \frac{K_{i,o}}{K_{i,o} + A}$$

**Marker**

$$R_{ac} = \frac{dA}{dt} + \frac{F_{in,s}}{W} A$$

### Oxidative growth on glucose and acetate

$$R_{ac} < 0 \quad \Rightarrow \quad \frac{d}{dt} \begin{bmatrix} X \\ S \\ A \\ P \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & 0 \\ 0 & -k_4 \\ k_5 & k_7 \end{bmatrix} \begin{bmatrix} \mu_1 \\ \mu_3 \end{bmatrix} - \frac{F}{V} \begin{bmatrix} X \\ S - S_{in} \\ A \\ P \end{bmatrix}$$

$$\mu_1 = q_{s,crit} / k_1$$

$$\mu_3 = q_{ac} / k_4$$

$$q_{ac} = q_{ac,max} \left( \frac{A}{K_A + A} \right) \left( \frac{K_{i,A}}{K_{i,A} + A} \right)$$

Fig. 2 Operational model of class bioprocesses

# Experimental data of fed-batch phytase production

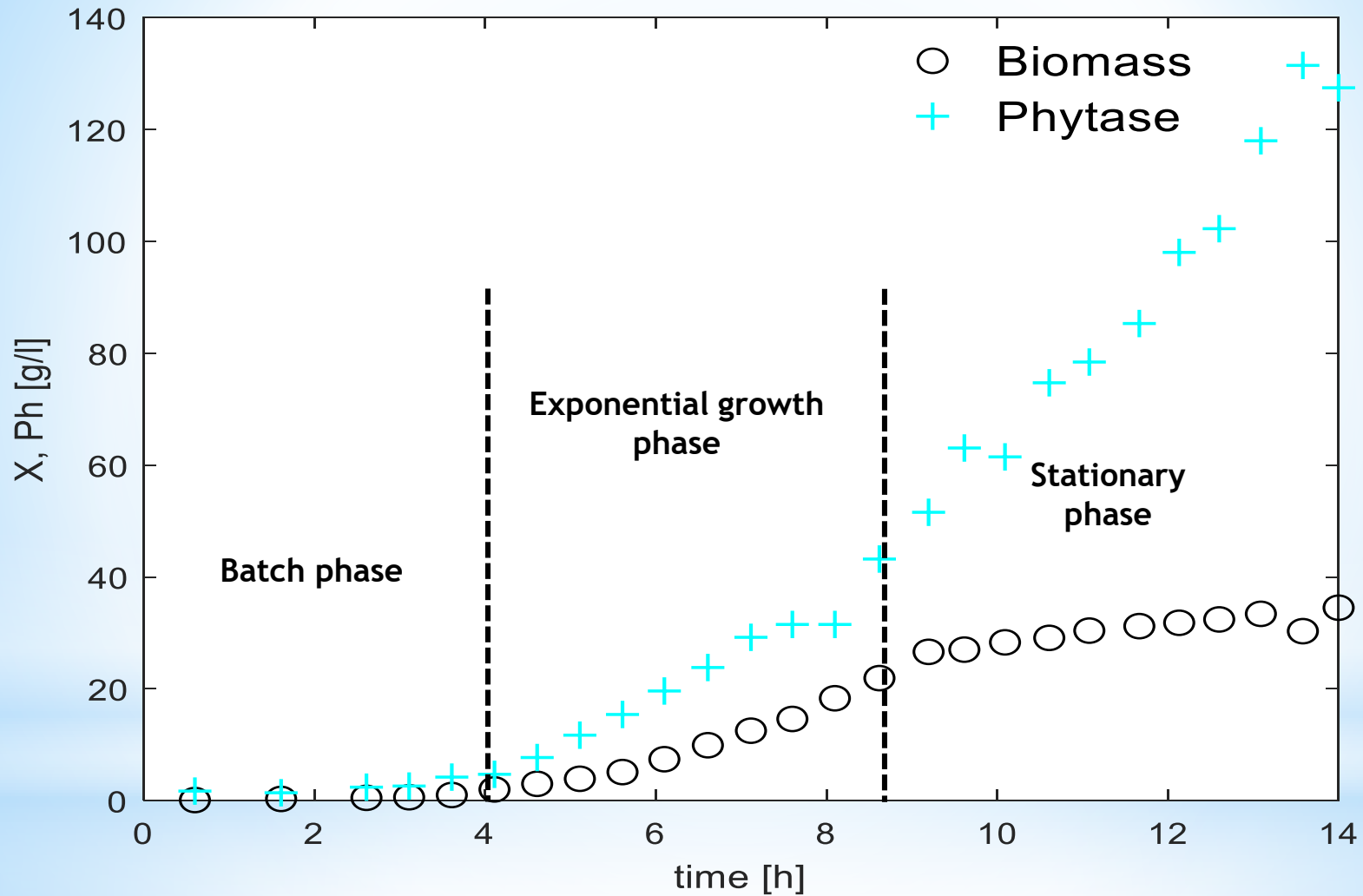


Fig. 3 Experimental data of biomass and phytase concentrations



# Experimental data of fed-batch phytase production

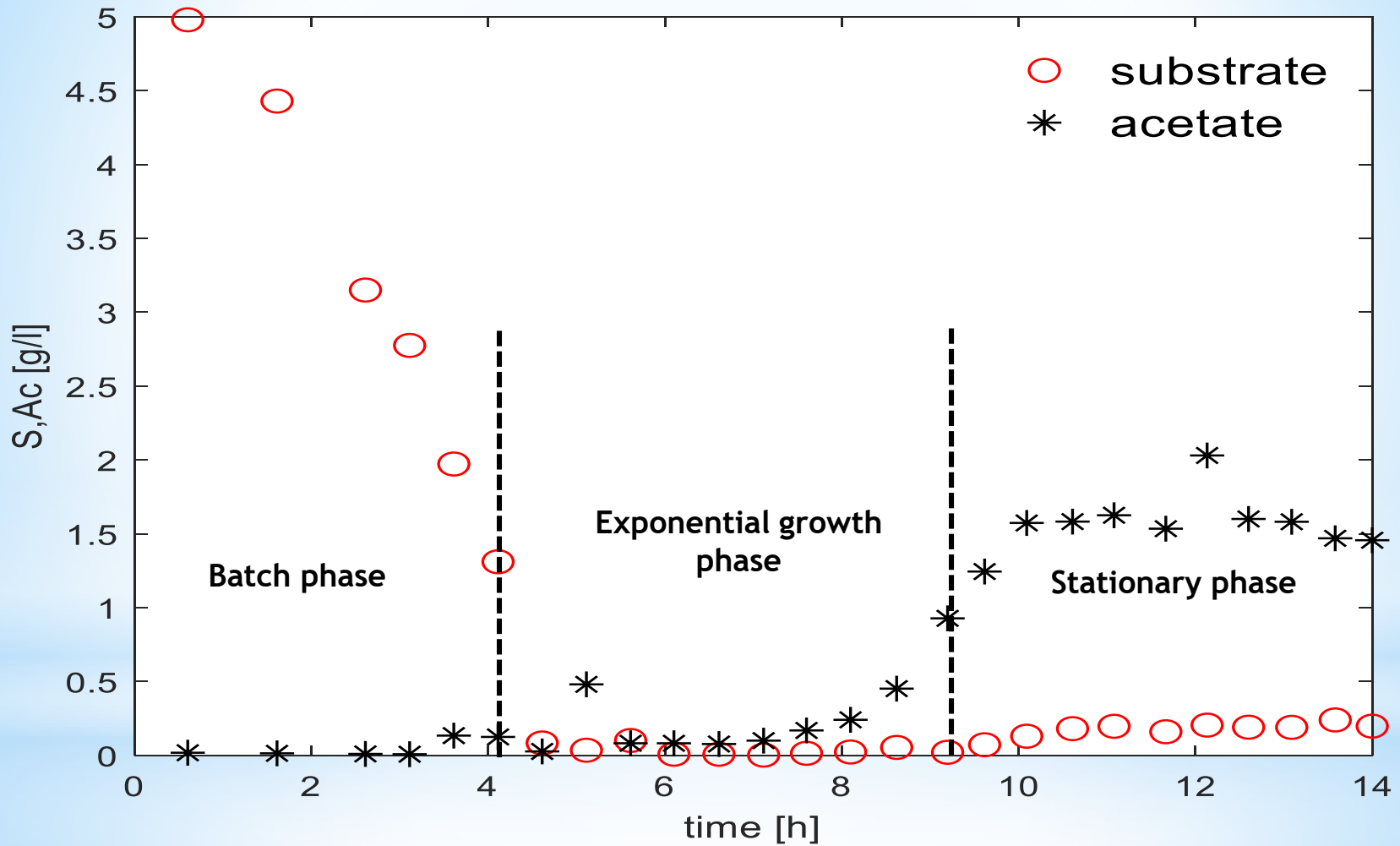


Fig. 4 Experimental data of substrate and acetate concentrations

# Results

Table 1. Estimated kinetic parameters

	$q_{lsmax}$	$k_s$	$k_{is}$	$q_{lomax}$	$k_{os}$	$k_{io}$	$q_{lacmax}$	$k_a$	$k_{ia}$	$k_1$	$k_2$	$k_3$	$k_4$	$k_5$	$k_6$	$k_7$
1 phase	4.19	0.19	5.54	1.1	2.15	0.088	0.082	1.17	-	3.69	0.557	0.19	4.6	1.41	2.66	0.45
2 phase	34.24	0.79	1.83	0.469	2.53	0.197	0.143	0.97	0.246	2.08	2.167	0.05	4.1	2.88	1.52	0.5
3 phase	77.11	0.47	12.3	2.1	3.29	0.134	0.002	0.295	0.228	16.6	11.66	0.42	9.9	39.45	9.53	0.56

# Simulation results

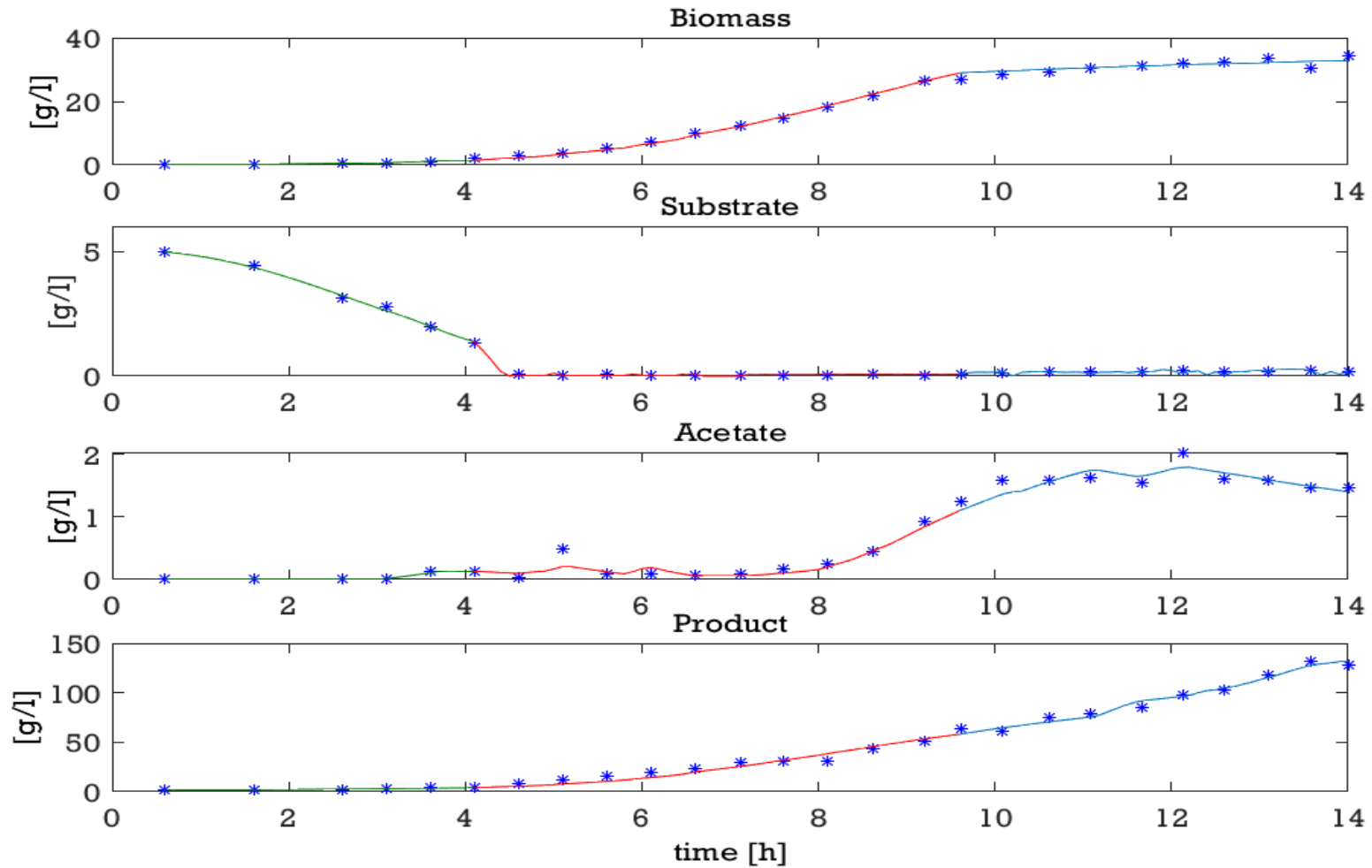


Fig. 5 Simulation results – models values of the biomass, substrate, acetate, and product concentrations are compared with experimental data for the three phases: batch phase – green lines, exponential growth phase – red lines, and stationary phase – blue lines.

# Simulation results

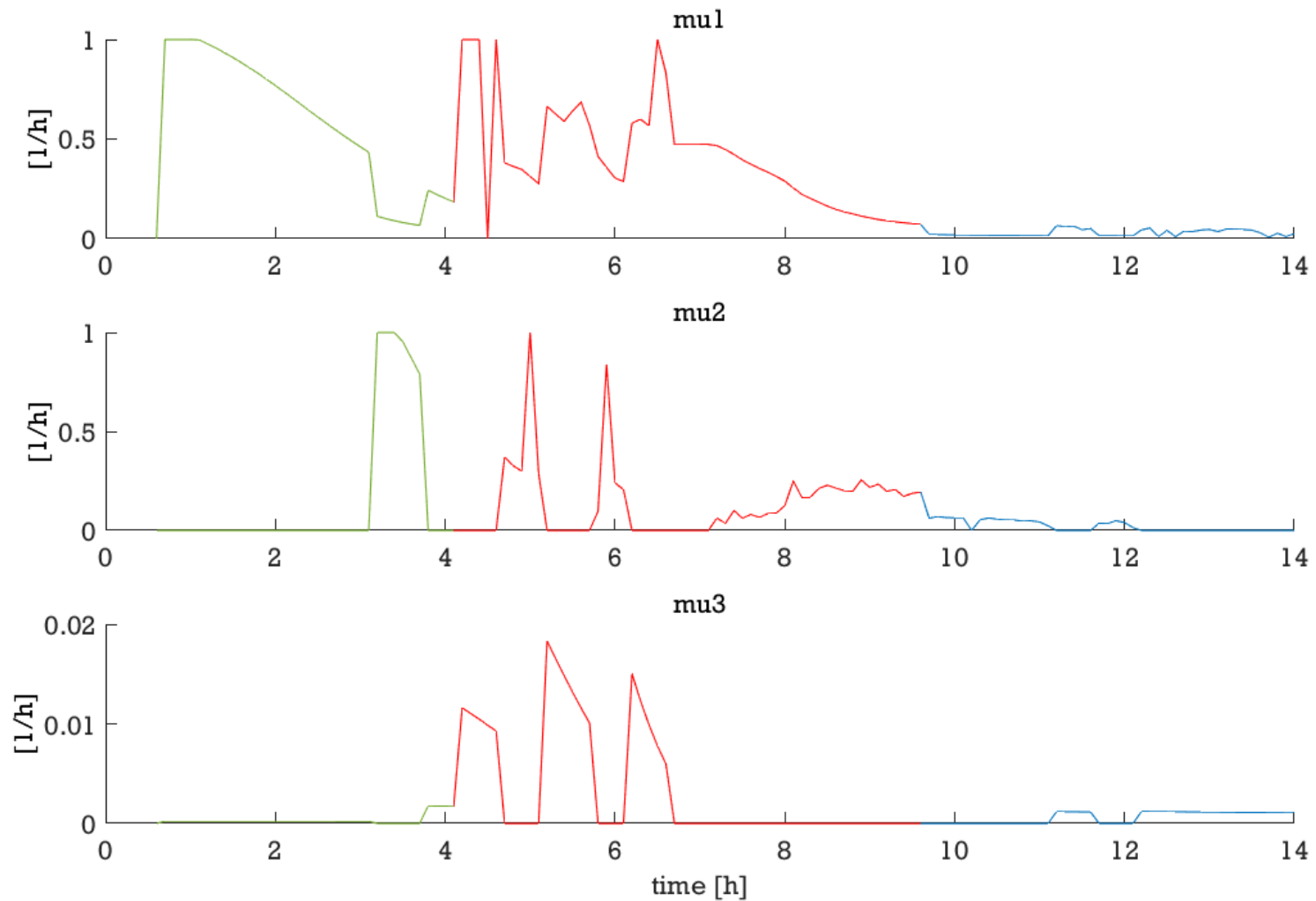


Fig. 6 Simulation results – models values of the three specific growth rates for the three phases: batch phase – green lines, exponential growth phase – red lines, and stationary phase – blue lines.

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen

## Biochemical model

$$\frac{dV}{dt} = F_{in}$$

$$\frac{d(S.V)}{dt} = -q_S(X.V) + F_{in}S_f$$

$$\frac{d(X.V)}{dt} = \mu(X.V)$$

$$\frac{d(A.V)}{dt} = (q_a^p - q_a^c)(X.V)$$

$$\frac{d(C_O.V)}{dt} = -q_O(X.V) + K_{La}(N).V.(C_O^* - C_O)$$

$$q_S = q_S^{\max} \frac{S}{k_S + S}$$

(1)

When  $q_S < q_S^{crit}$

$$\mu = q_S Y_{SX}^{oxid}$$

$$q_a^p = 0$$

$$q_a^c = \min \left( \frac{q_a^{c, \max} A}{k_a + A}, \frac{q_O^{\max} - q_S Y_{OG}}{Y_{Oa}} \right)$$

$$q_O = q_S Y_{OG}$$

When  $q_S \geq q_S^{crit}$

$$\mu = q_S^{crit} Y_{SX}^{oxid} + (q_S - q_S^{crit}) Y_{SX}^{ferm}$$

$$q_a^p = (q_S - q_S^{crit}) Y_{SA}$$

$$q_a^c = 0$$

$$q_O = q_S^{crit} Y_{OG}$$

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen

**Operational model** when  $q_s < q_s^{crit}$

$$(2) \quad \begin{aligned} \frac{dV}{dt} &= F_{in}; \\ \frac{d(S.V)}{dt} &= -\varphi_1 + F_{in} S_f; \\ \frac{d(X.V)}{dt} &= k_1 \varphi_1; \\ \frac{d(C_o.V)}{dt} &= -k_3 \varphi_1 + Q_{in}.V; \end{aligned} \quad \text{where} \quad \begin{aligned} k_1 &= Y_{SX}^{oxid}, \\ k_3 &= Y_{OG}; \\ \varphi_1 &= q_s V . X; \\ Q_{in}.V &= K_{La} (N)(C_o^* - C_o).V; \end{aligned}$$

Operational model when  $q_s < q_{s,crit}$

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen

**Operational model**

when  $q_S \geq q_S^{crit}$

$$\frac{dV}{dt} = F_{in};$$

$$\frac{d(S.V)}{dt} = -\varphi_1 - \varphi_2 + F_{in} S_f;$$

$$\frac{d(X.V)}{dt} = k_1 \varphi_1 + k_2 \varphi_2;$$

$$\frac{d(C_o.V)}{dt} = -k_3 \varphi_1 + Q_{in}.V;$$

$$\frac{d(OUR.V)}{dt} = k_4 \varphi_1 + k_5 \varphi_2.$$

where

$$k_1 = Y_{SX}^{oxid},$$

$$k_2 = Y_{SX}^{fe}$$

$$k_3 = Y_{OG};$$

$$k_4 = k_1 k_3 q_S^{crit}$$

$$k_5 = k_2 k_3 q_S^{crit}$$

$$\varphi_1 = q_S^{crit} V.X;$$

$$\varphi_2 = (q_S - q_S^{crit}) V.X;$$

$$OUR.V = k_3 q_S^{crit} X.V;$$

$$Q_{in}.V = K_{La}(N)(C_o^* - C_o).V;$$

(3)

Operational model when  $q_S \geq q_{S,crit}$

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen

$$\frac{dZ_1}{dt} = -\frac{1}{k_3} Q_{in} \cdot V + F_{in} S_f \quad Z_1 = -\frac{1}{k_3} C_o \cdot V + S \cdot V$$

$$\frac{dZ_2}{dt} = \frac{k_1}{k_3} Q_{in} \cdot V \quad Z_2 = \frac{k_1}{k_3} C_o \cdot V + X \cdot V$$

**Adaptive Biomass Observer for the case  $q_S < q_S^{crit}$**

$$\frac{dV}{dt} = F_{in}$$

$$\frac{dZ_1}{dt} = -\frac{1}{k_3} Q_{in} \cdot V + F_{in} S_f \quad (4)$$

$$\frac{dZ_2}{dt} = \frac{k_1}{k_3} Q_{in} \cdot V$$

$$\hat{X} = (Z_2 + k_1 Z_1) / V$$



# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen

$$Z_1 = -\frac{1}{k_3} C_o V - \frac{1}{k_3} OUR.V + S.V$$

$$\frac{dZ_1}{dt} = -\frac{1}{k_3} Q_{in}.V + F_{in} S_f$$

$$Z_2 = \frac{k_1 q_S^{crit} - k_1}{k_2 k_3 q_S^{crit}} C_o V + \frac{k_1 q_S^{crit} - k_2}{k_2 k_3 q_S^{crit}} OUR.V + XV$$

$$\frac{dZ_2}{dt} = \frac{k_1 q_S^{crit} - k_1}{k_2 \cdot k_3 q_S^{crit}} Q_{in}.V$$

**Adaptive Biomass Observer for the case  $q_S \geq q_S^{crit}$**

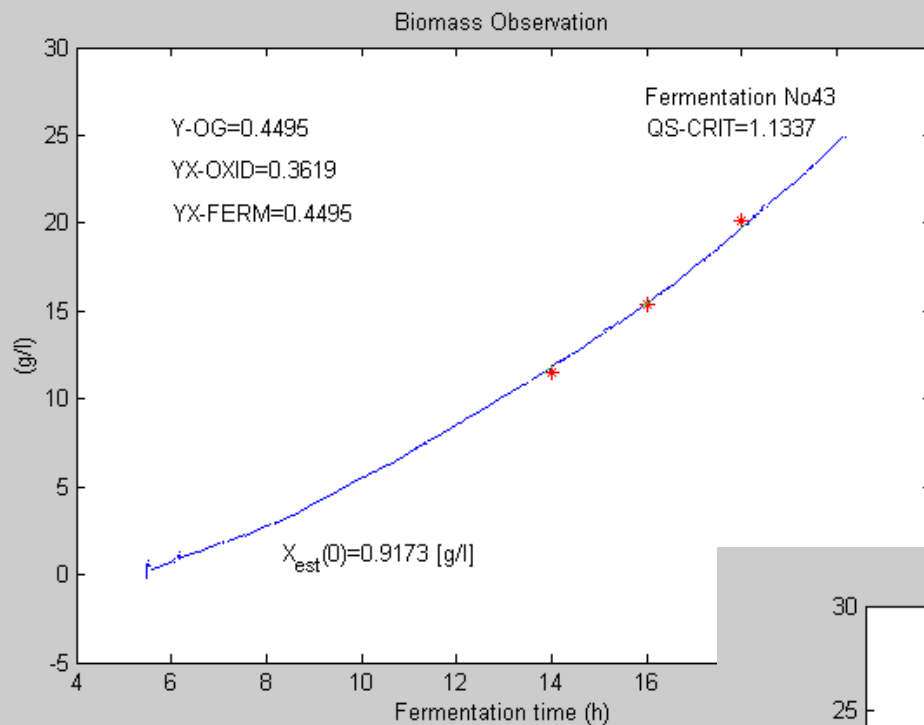
$$\frac{dV}{dt} = F_{in}$$

$$\frac{dZ_1}{dt} = -\frac{1}{k_3} Q_{in}.V + F_{in} S_f \quad (5)$$

$$\frac{dZ_2}{dt} = \frac{k_1 q_S^{crit} - k_1}{k_2 k_3 q_S^{crit}} Q_{in}.V$$

$$\hat{X} = \left( Z_2 + \frac{k_1 q_S^{crit} - k_1}{k_2 q_S^{crit}} \left( Z_1 + \frac{1}{k_3} OURV \right) + \frac{k_1 q_S^{crit} - k_2}{k_2 q_S^{crit}} \left( Z_1 + \frac{1}{k_3} C_o.V \right) \right) / V$$

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen



(a)

$$q_S \geq q_S^{crit}$$

$$q_S < q_S^{crit}$$

(b)

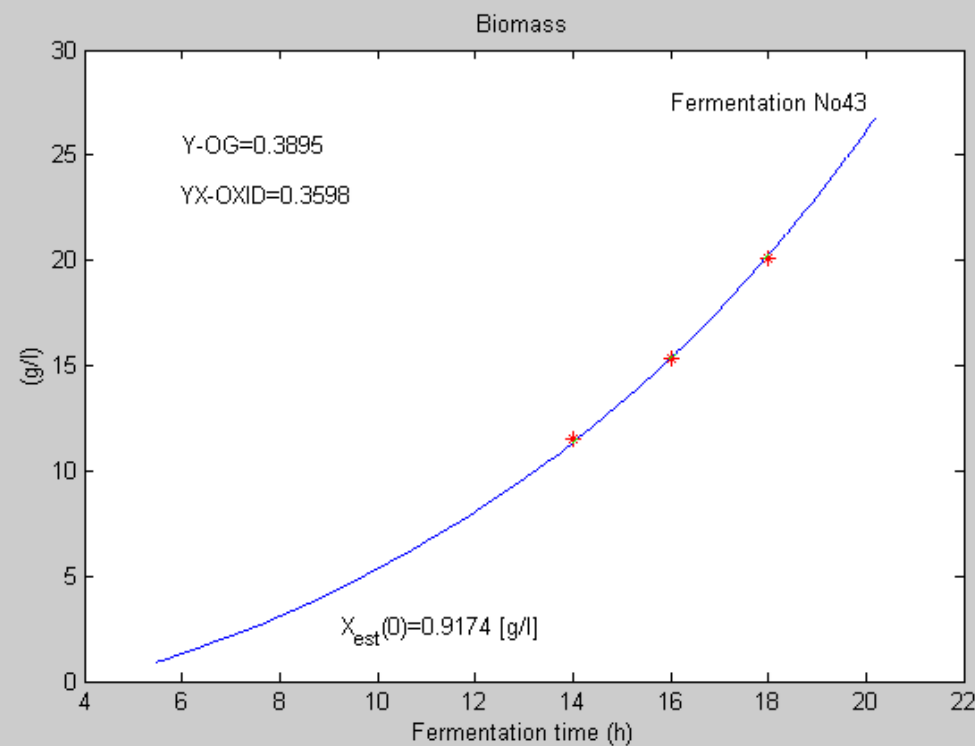
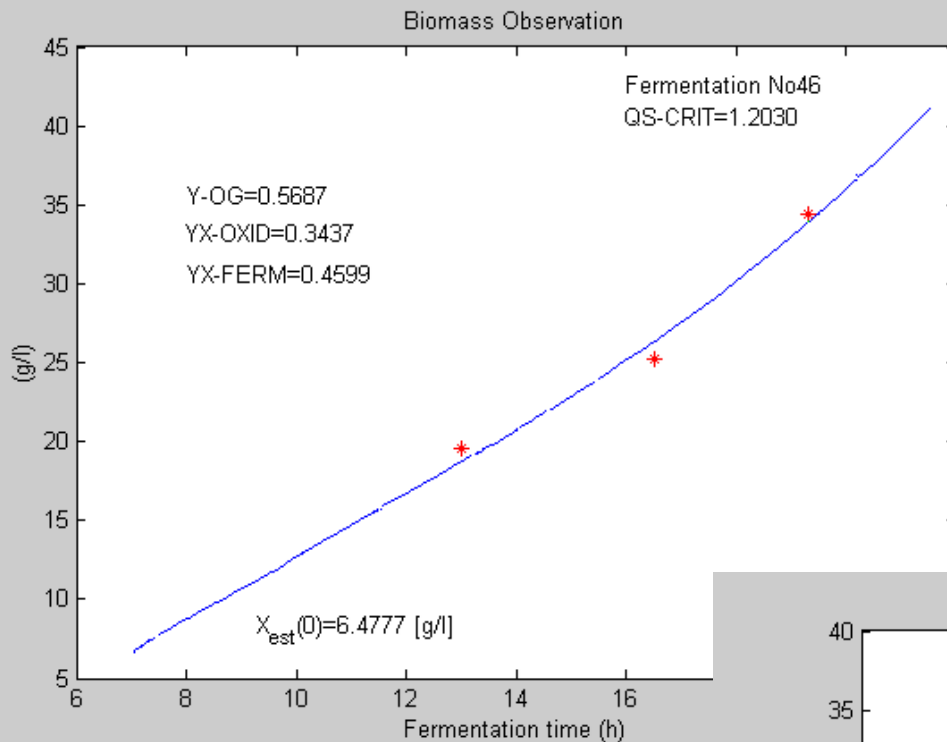


Figure 7

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen



$$q_S \geq q_S^{crit}$$

(a)

$$q_S < q_S^{crit}$$

(b)

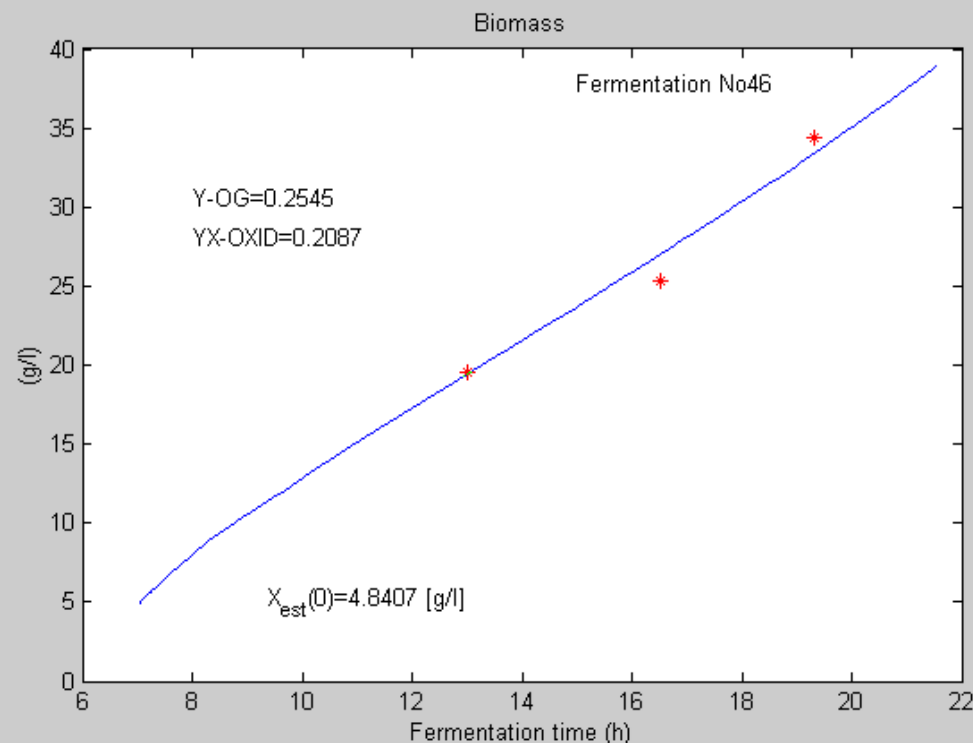
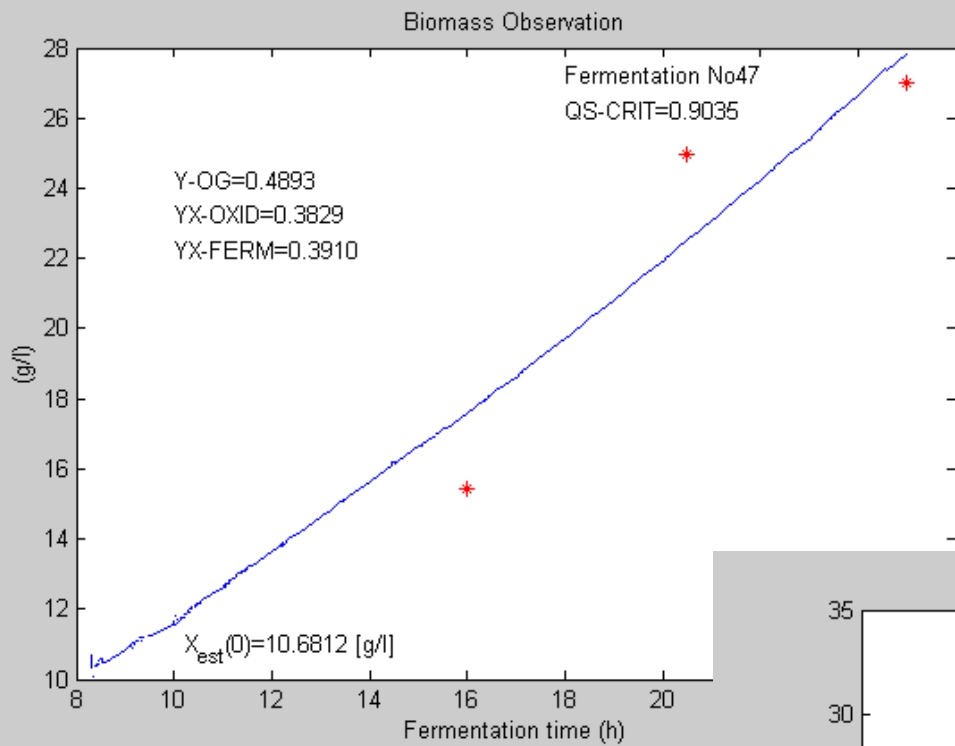


Figure 8

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen



$$q_S \geq q_S^{crit}$$

(a)

$$q_S < q_S^{crit}$$

(b)

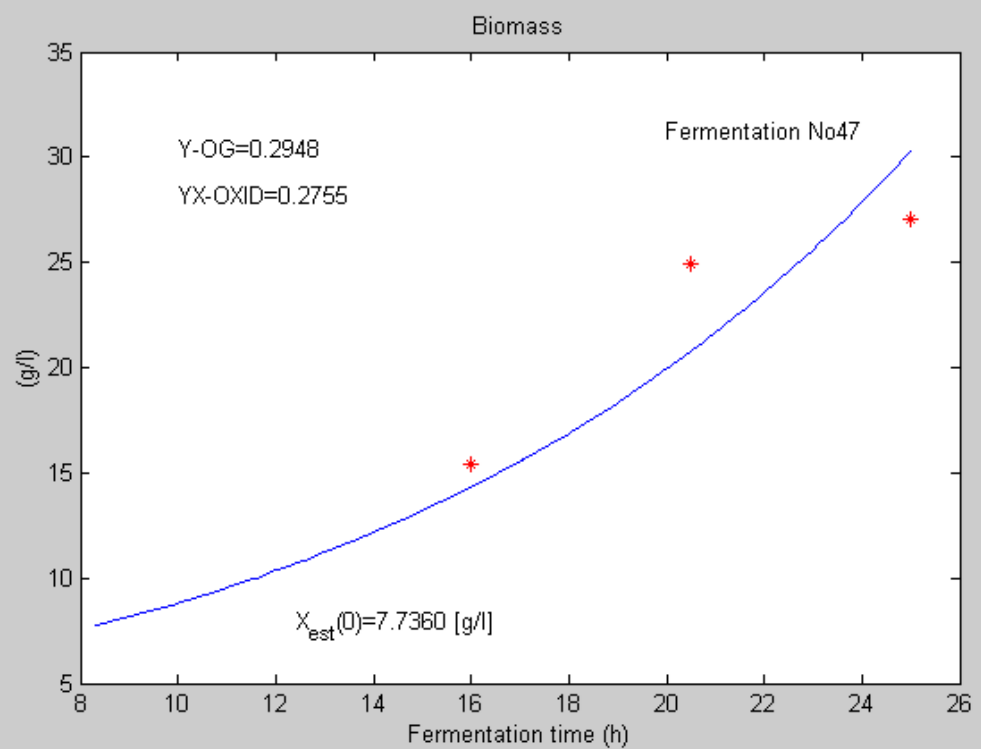
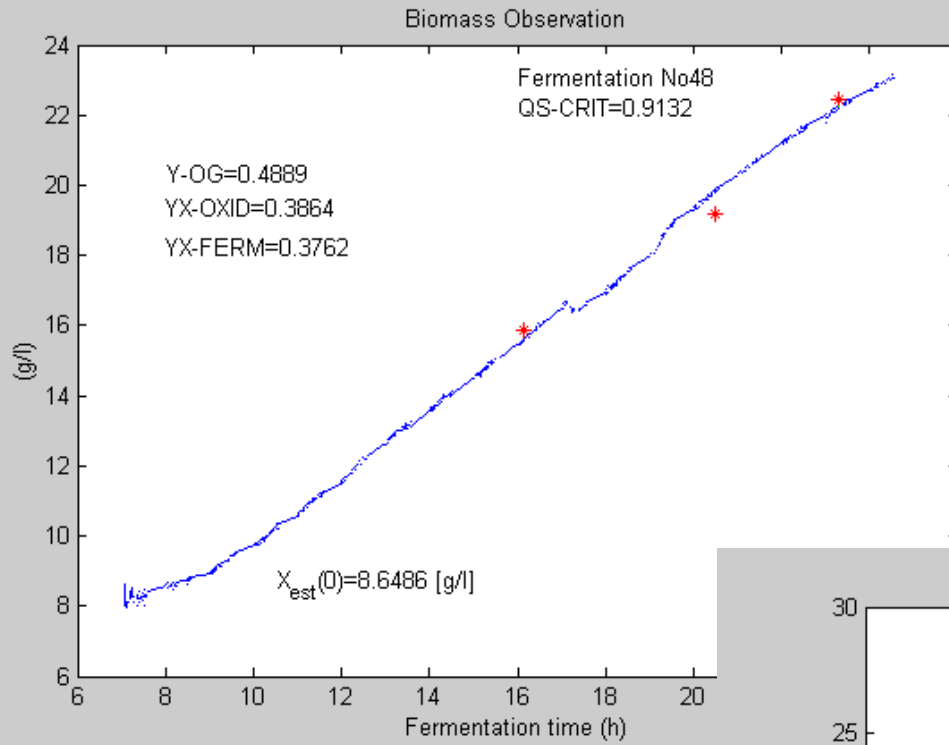


Figure 9

# Adaptive Biomass Observer in Fed-batch Cultivation of *Escherichia coli* on the Basis of On-line Measurements of Oxygen



$$q_s \geq q_s^{crit}$$

(a)

$$q_s < q_s^{crit}$$

(b)

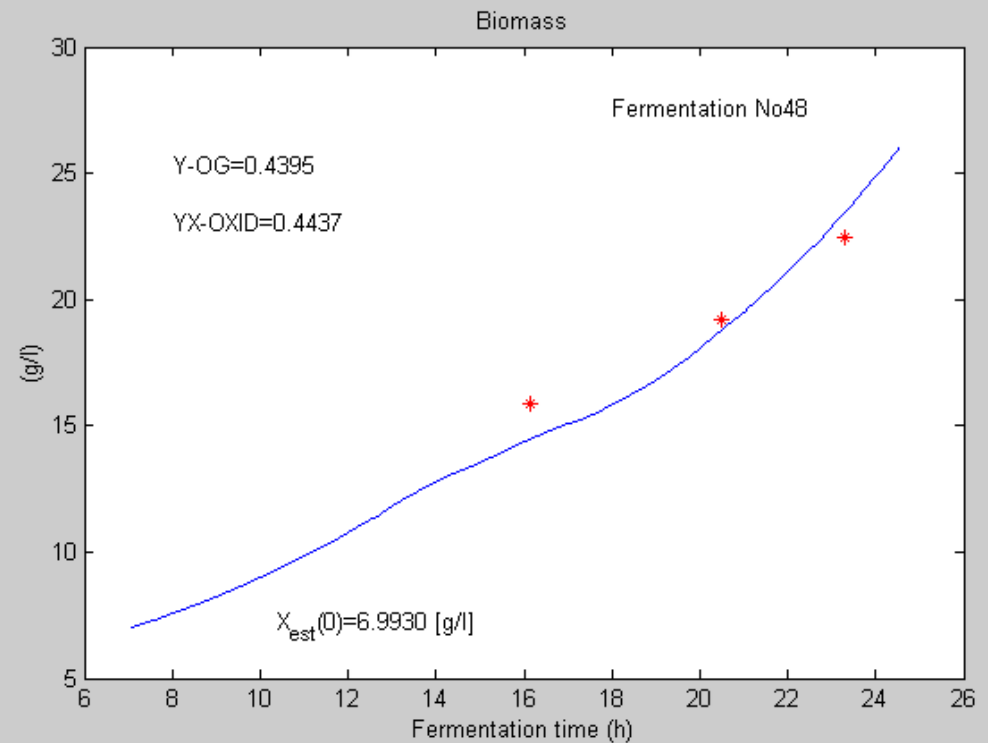


Figure 10

# Adaptive Control of Protein Production Bioprocess with Three Physiological States

Operational process model describing three physiological states

$$\frac{d}{dt} \begin{bmatrix} X \\ S \\ A \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & -k_2 \\ 0 & k_3 \end{bmatrix} \begin{bmatrix} \mu_1(t) \\ \mu_2(t) \end{bmatrix} X - D \begin{bmatrix} X \\ S \\ A \end{bmatrix} + \frac{F_{in,S}}{W} \begin{bmatrix} 0 \\ S_{in} \\ 0 \end{bmatrix} \quad (6)$$

$$\frac{d}{dt} \begin{bmatrix} X \\ S \\ A \end{bmatrix} = \begin{bmatrix} 1 & 1 \\ -k_1 & 0 \\ 0 & -k_4 \end{bmatrix} \begin{bmatrix} \mu_1(t) \\ \mu_3(t) \end{bmatrix} X - D \begin{bmatrix} X \\ S \\ A \end{bmatrix} + \frac{F_{in,S}}{W} \begin{bmatrix} 0 \\ S_{in} \\ 0 \end{bmatrix} \quad R_a = \frac{dA}{dt} + \frac{F_{in,S}}{W} A$$

$R_a = 0$  oxidative growth on glucose

$R_a > 0$  oxidative-fermentative growth on glucose

$R_a < 0$  oxidative growth on acetate

# Adaptive Control of Protein Production Bioprocess with Three Physiological States

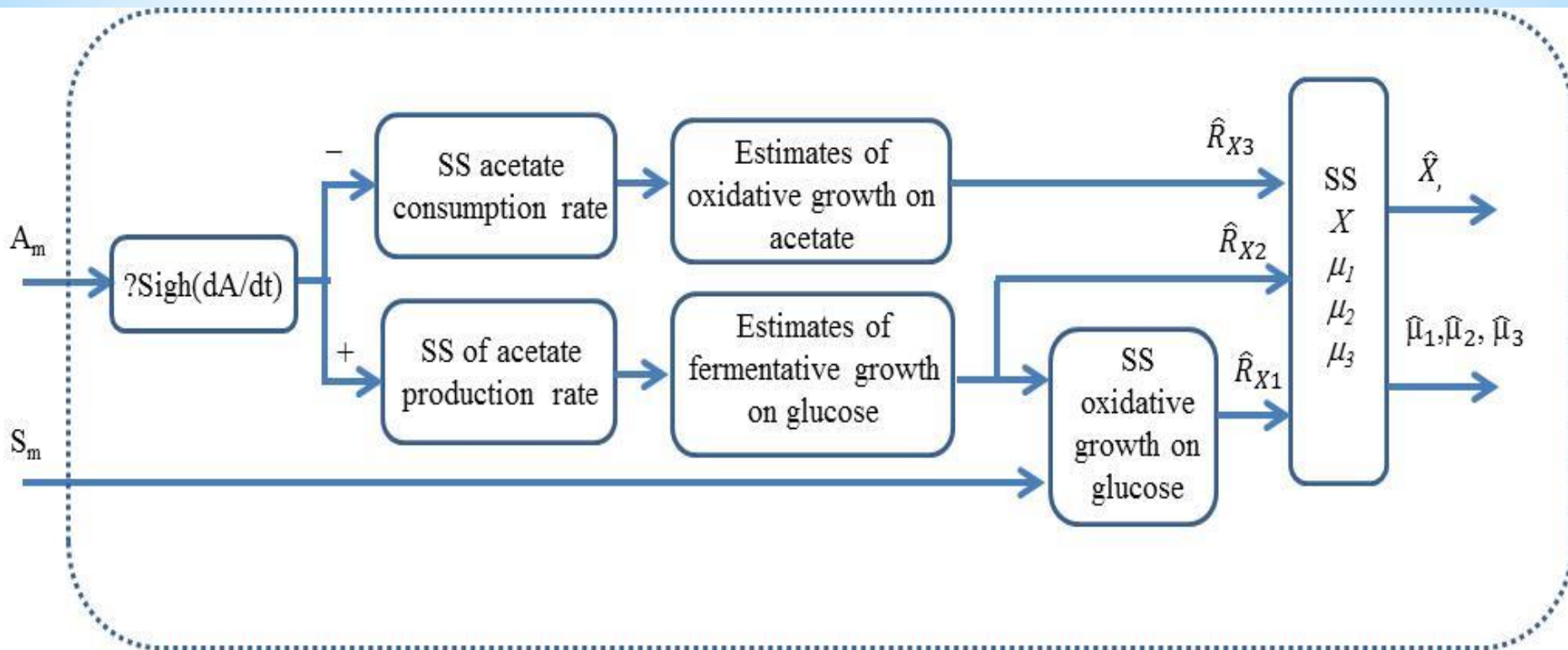


Figure 11 Cascade structure of the software sensor for monitoring of three metabolic states

# Adaptive Control of Protein Production Bioprocess with Three Physiological States

Oxidative growth  
on glucose



$$\frac{d\hat{S}}{dt} = -k_1\hat{R}_{X1} - k_2\hat{R}_{X2} - DS_m + \frac{F_{in,S}}{W}S_{in} + w_3(S_m - \hat{S})$$

$$\frac{d\hat{R}_{X1}}{dt} = w_4(S_m - \hat{S})$$

Oxidative-fermentative  
growth on glucose



$$\frac{d\hat{A}}{dt} = \hat{R}_{ap} - DA_m + w_1(A_m - \hat{A})$$

$$\frac{d\hat{R}_{ap}}{dt} = w_2(A_m - \hat{A})$$

$$\hat{R}_{X2} = \hat{R}_{ap}/k_3$$

$$\frac{d\hat{S}}{dt} = -k_1\hat{R}_{X1} - k_2\hat{R}_{X2} - DS_m + \frac{F_{in,S}}{W}S_{in} + w_3(S_m - \hat{S})$$

$$\frac{d\hat{R}_{X1}}{dt} = w_4(S_m - \hat{S})$$

$$\frac{d\hat{X}}{dt} = \hat{R}_{X1} + \hat{R}_{X2} - D\hat{X}$$

$$\hat{\mu}_1 = \hat{R}_{X1}/\hat{X}$$

$$\hat{\mu}_2 = \hat{R}_{X2}/\hat{X}$$

$$\frac{d\hat{S}}{dt} = \hat{R}_S - DS + \frac{F_{in,S}}{W}S_{in} + w_5(S_m - \hat{S})$$

$$\frac{d\hat{R}_S}{dt} = w_6(S_m - \hat{S})$$

Oxidative growth  
on acetate



$$\frac{d\hat{A}}{dt} = \hat{R}_{ac} - DA + w_5(A - \hat{A})$$

$$\frac{d\hat{R}_{ac}}{dt} = w_6(A - \hat{A})$$

$$\hat{R}_{X3} = -\hat{R}_{ac}/k_4$$

$$\frac{d\hat{X}}{dt} = \hat{R}_{X1} + \hat{R}_{X2} + \hat{R}_{X3} - D\hat{X}$$

$$\hat{\mu}_3 = \hat{R}_{X3}/\hat{X}$$

$$\hat{\mu}_1 = \hat{R}_{X1}/\hat{X}$$

$$\hat{\mu}_2 = \hat{R}_{X2}/\hat{X}$$

Figure 12 Observer based estimators grouped, depending on the physiological state of the process



## Adaptive Control and Results

$$F = \frac{W \cdot (-\lambda(S^* - S_m) + k_1 \hat{R}_{X1} + k_2 \hat{R}_{X2} + k_4 \hat{R}_{X3})}{S_{in} - S_m} \quad (7)$$

# Adaptive Control of Protein Production Bioprocess with Three Physiological States

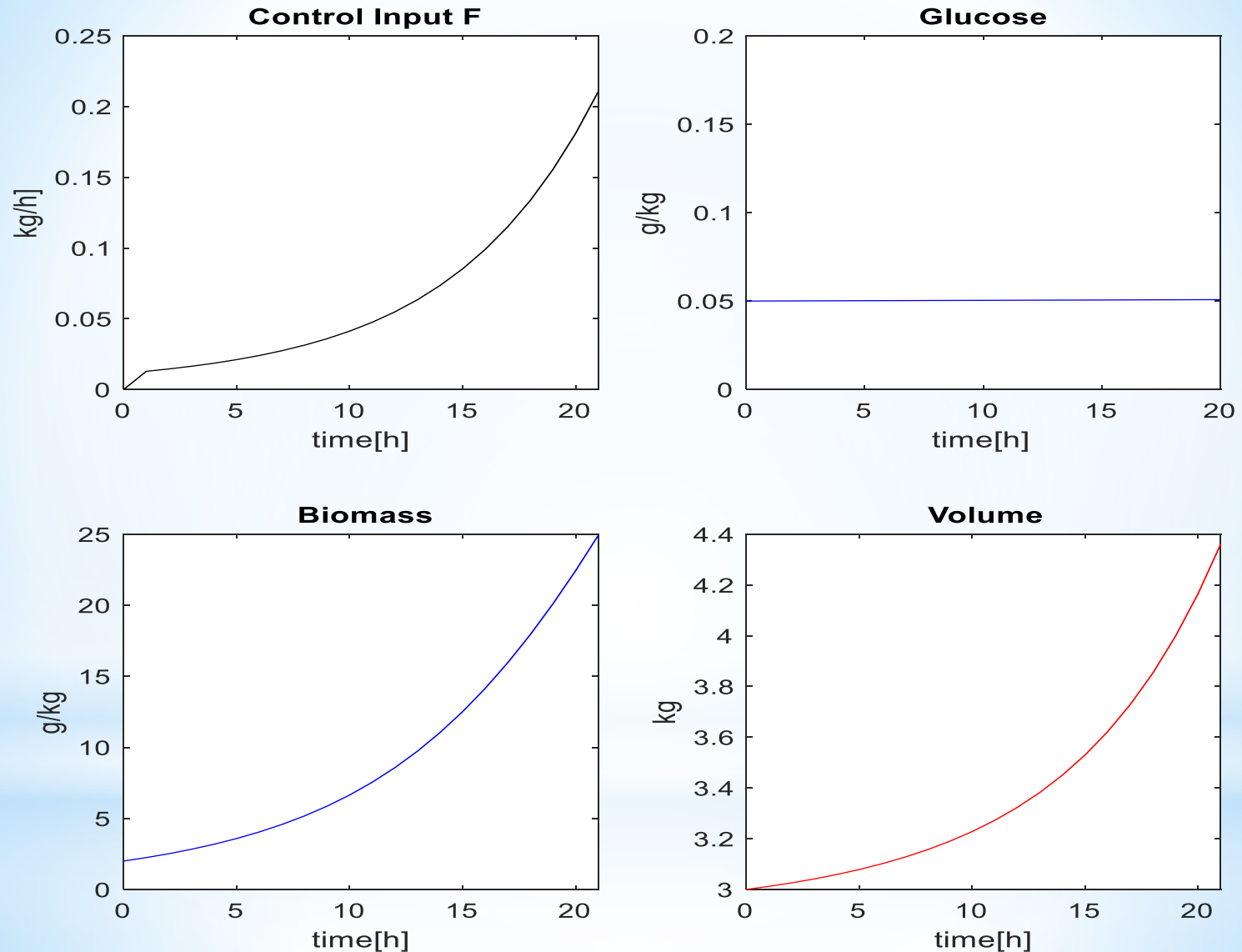


Figure 13 Linearizing control algorithm investigation

# Adaptive Control of Protein Production Bioprocess with Three Physiological States

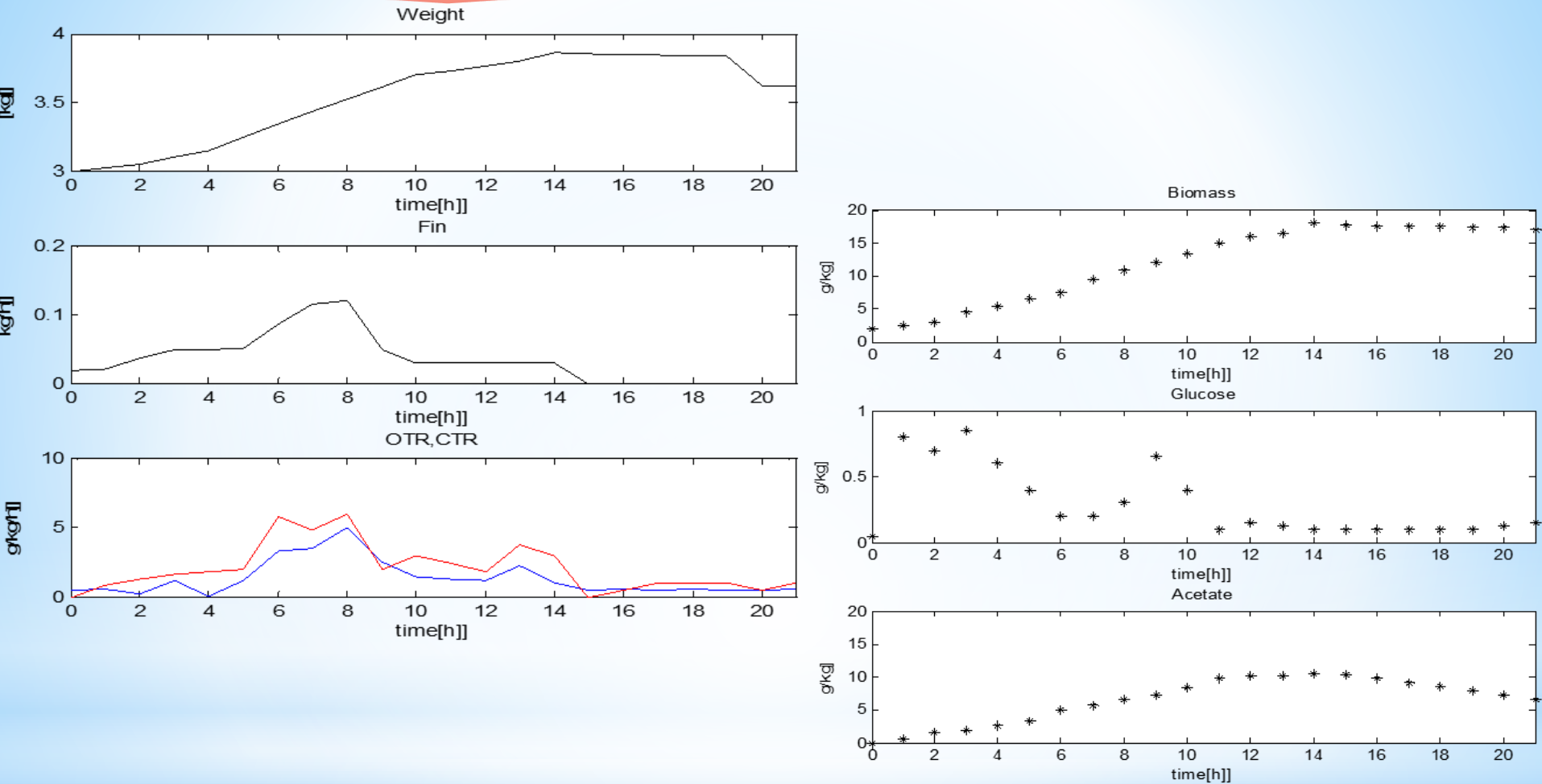


Figure 14 Open loop control for the fermentation according the experimental data

# Further research and work on the system

Two modules have been planned initially as part of the interactive system InSEMCoBio.

The module for model parameter identification of the system is currently under development. It will be further expanded with new hybrid metaheuristic algorithms and different models of cultivation processes.

The work on the system InSEMCoBio will continue further by developing the second module for an adaptive control design

## Acknowledgements

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**THANKS FOR YOUR  
ATTENTION**